



Nigerian Journal of Soil Science

Journal homepage: www.soiljournalnigeria.com

Effect of simulated acid rain on growth and micronutrient uptake of okra (*Abelmoschus esculentus*) on coastal plain sands in Abia State, South-eastern, Nigeria

Eteng, E.U¹. and Essien, G. E².

1. Department of Soil Science and Meteorology, Michael Okpara University of Agriculture, Umudike Nigeria

2. Department of Soil Science and Land Management Resources, University of Uyo, Uyo Nigeria

ARTICLE INFO

Article history:

Received 28th March 27, 2020

Received in revised form April 7, 2020

Accepted April 28, 2020

Available online May 23, 2020

Keywords:

Acid rainfall

Frequency of application

Okra

Micronutrients consumption

ABSTRACT

The study was conducted in April, 2019 at Umudike to examine the impact of simulated acid rain (SAR) and frequency of SAR spray on growth and micronutrients content of okra (*Abelmoschus esculentus* (L.) Moench) in an ultisol at Umudike. The treatments consisted of three (3) pH levels: Coca-cola solution (pH 2.7) and SAR (pH 3.5 and 5.5) as well as control (water) and; three (3) frequency of foliar spray at 1, 2 and 3 day weekly, arranged according to their respective pH values and spray treatments in a completely randomized design (CRD) replicated three times. Plant leaves were analysis for micronutrients (Cu, Fe, Mn and Zn) content. The result shows that, under the stress of SAR and frequency of SAR application, growth parameters measured; leaf number, shoot height and leaf area were significantly reduced, compared to the control treatment. Nutritional analysis of the leaves also revealed that, the content micronutrients in okra, increased with declining pH value of AR and frequency of application, from pH 5.5 to pH 2.7. Coke solution (pH 2.7) and three (3) days per weekly application of SAR gave the highest reduction in plant growth parameter in Okra but, had higher increased in micronutrients content, respectively. Further result showed that okra (*Abelmoschus esculentus* (L.) Moench) is resistant to SAR from pH ≥ 5.5 in terms of growth and nutritional values.

Corresponding Author's E-mail Address:

eteng_em@yahoo.com .+2347030882864

ISSN-1597-4488 ©publishingrealtime.

All right reserved.

1.0. Introduction

Acid rainfall (AR) is a form of wet deposition that has been acidified when pollutants, such as oxides of sulphur and nitrogen, contained in power plant emission, factory smoke and car exhaust, react with the moisture in the atmosphere (Kita *et al.*, 2004), with elevated levels of hydrogen ions (low pH) (Prashant, 2010). The processes of acid rains (AR) and other types of acidic deposition, such as acid snow, hail, dew, and fog, form when the emissions of SO₂ and NO, react with water molecules in the atmosphere to produce acids (Chesapeake, 2013). These pollutants (SO₂, NO_x), once released into the atmosphere, can be converted chemically (under the influence of sunlight and moisture) into secondary pollutants such as sulphuric

acid (H₂SO₄) and nitric acid (HNO₃), which fall as acid rain (Li and Gao, 2002).

Plants exposure to Acid rainfall (AR) results in characteristic foliar injury symptoms, modified leaf anatomy, structural changes in the photosynthetic pigment apparatus and a decrease in chlorophyll a and b contents (Sant' Anna-Santos *et al.*, 2006). Exposure to simulated acid rain (SAR) alters the ability of plants to take in CO₂ for photosynthesis and consequently inhibits the production of glucose (Sunita and Sharad, 2013). Singh and Agrawal (2001) noted that the shoot and root lengths, leaf areas, and total biomass or yield of two varieties of wheat declined signifi-

cantly at or below pH 4.0 and concluded that SAR had a significantly negative effect on wheat. Compared with other crops such as cereals, root and tubers, *etc.*, vegetables crops are expected to be more sensitive to AR (Meng *et al.*, 2011). Based on the field investigations and experimental results, the effects of AR on vegetables include poor seed germination and seedling growth, leaf injury (Meng *et al.*, 2011), interference with normal metabolism (Johnston *et al.*, 1986; Khan and Shikha, 2004), growth and yield (Dubay and Allen, 1987) as well as nutritional quality (Nduka, *et al.*, 2008). There are several mechanisms by which AR influences plant growth, as by direct foliar injury or indirect by affecting root's soil-water relations (Prashant, 2010; Chesapeake, 2013).

Okra is a very sensitive crop to high concentrations of acid levels like many other vegetable plants. Foliar applications of SAR have been shown to result in significant reductions in *Abelmoschus esculentus* yield and quality (Eguagie *et al.*, 2016). *Abelmoschus esculentus* was therefore chosen for this experiment since, Lee *et al.* (1981) demonstrated an increase in its productivity with acidic rainfall with a pH <4.5, while Eguagie *et al.*, (2016) under greenhouse and agronomic field conditions, showed that, foliar injury was accompanied by a decline in growth and yield due to low pH levels of simulated acid rain (SAR <4.5). However, field studies reported by Odiyi and Bamidele (2014), Odiyi and Eniola (2014), and Milton and Abigael (2015) revealed that simulated AR caused reduction in plant growth and yield of cassava, cowpea and okra, respectively. This more or less supports the preceding reports.

The adverse effects of acid rain which include; chlorosis, necrosis, early senescence and stunting have been reported by Kausar *et al.*, (2005) and Bamidele and Eguagie, (2015). Also, Exposure of plants to AR which resulted in characteristic foliar injury symptoms, modified leaf anatomy, structural changes in the photosynthetic pigment apparatus and a decrease in the chlorophyll concentrations were reported by Chesapeake, (2013), Stoyhora and Velikova, (2004), Sant' Anna-Santos *et al.*, (2006).

Acid rain deposition have also been reported to cause a reduction in soil fertility as well as leaching of metals which may lead to depletion of essential nutrient minerals from leaves and soil (Butnariu and Samfira, 2013; Eguagie *et al.*, 2016). With improper functioning of leaves, the growth of tree crops is retarded, resulting in loss of leaves and stunted growth.

Little or no information is currently available concerning the effects of simulated acid rainfall and frequency of foliar spray on growth and nutrient content of Okra in Umudike, South-eastern Nigeria. The study therefore, examined the impact of SAR and frequency of foliar spray on growth performance and micronutrient content of okra (*Abelmoschus esculentus* (L.) Moench) on a coastal plain sands soil in south-eastern Nigeria.

2.0. Materials and methods

2.1. Description of the study area

The study was conducted in a greenhouse environment from April to September, 2019 in Michael Okpara University of

Agriculture, Umudike, Abia State, Nigeria (Latitude 05° 48' 21' N and Longitude 07° 53' 94' E), with an altitude of 116 meter above sea level., from the site is level to gentle sloping. The study area falls within the tropical rain forest with mean annual rainfall of 2200 mm, distributed over 9 months in bimodal rainfall pattern, with 5 months of dry season and short dry period popularly called August break. The relative humidity varies from 84% to 87% while, monthly minimum air temperature ranged from 20°C to 24°C and the monthly maximum air temperature ranged from 28°C to 35°C (NRCRI, 2018).

2.2. Preparation of simulated acid rain (SAR)

The simulated acid rain was formed from a mixture of concentrated sulphuric acid (H₂SO₄) and concentrated nitric acid (HNO₃) in a ratio of 3:1. A 60 ml of sulphuric acid and 20 ml of nitric acid were measured with the measuring cylinder into a conical flask containing distilled water and was made up to 2 liters. The acidic solutions were then calibrated using concentrated NH₄OAc solution and distilled water with a Deluxe pH meter to get the desired pH (3.5. and 5.5), Coca-cola solution contains phosphoric acid with a pH of 2.7 (Eteng *et al.*, 2014) and cross checked with pH meter. There was a control which comprised no AR.

2.3. Treatments and experimental design

Okra seeds (Var. Clemsor spineless) produced by Premier Seeds in Nigeria, were obtained from National Seed Council Umudike, Abia State, Nigeria. The soil used for the experiment was obtained from from the 0-20 cm soil depth in the Michael Okpara University of Agriculture Farm.

Four (4) levels of treatments consisted of three (3) pH levels: one Coca-cola solution (pH 2.7) and two SAR (pH 3.5, 5.5) as well as a control (no treatment) and three (3) frequency of SAR application applied at 1, 2 and 3 times per week. A total of twelve (12) treatment combinations, arranged according to a completely randomized design (CRD) replicated three times.

2.4. Planting and foliar application of SAR procedures

Before planting, the okra seeds (Var. Clemsor spineless) were soaked for 12 hours in order to soften the seed coat and hasten germination. Three viable seeds of *Abelmoschus esculentus* were sown in each plastic pot and germination was achieved after 3 days of planting. Thinning was done after 14 days of planting, leaving two plants per pot of 10 liters capacity. The plants were thereafter grown for three weeks before the acid treatments commenced.

After three (3) weeks of planting, 10 ml each of SAR and Coca-cola solutions were foliar sprayed on the okra leaves at a frequency of once (1), twice (2) and thrice (3) weekly according to their pH values, respectively, using a hand-held medium size pressurized sprayer to mimic natural rainfall. The acid solution application was carried out fortnightly following the methods of Lal and Singh (2012) and Milton and Abigael, (2015). Th study lasted for eight weeks before the experiment was terminated.

2.5. Laboratory analysis

100g of the soil samples collected and used for the were air-

dried, sieved with a 2 mm sieve and stored for some chemical analysis. The soil chemical properties were analyzed according to standard procedures using the methods of IITA, 1979 and a modified version of Udo *et al.* (2009). The initial soil properties used for the pot experiment showed that, the soil was sandy loam, strongly acidic with pH of 5.16. Organic carbon was 7.14 g kg^{-1} with total N of 1.34 g kg^{-1} and available P of 6.12 mg kg^{-1} . Exchangeable bases were 2.25, 1.37, 0.26 and $0.14 \text{ cmol kg}^{-1}$ for Ca, Mg, K and Na, respectively. The values of the available micronutrients determined by AAS were, 0.8, 6.8, 5.40 and 2.00 mg kg^{-1} for Cu, Fe, Mn and Zn, respectively.

The extraction of available soil Cu, Fe, Mn and Zn and their nutritional values in okra plants were determined according to standard procedures using the atomic absorption spectrophotometer (AAS), as described by Meng *et al.* (2011). The values of the available micronutrients determined were, 0.8, 6.8, 5.40 and 2.00 mg kg^{-1} for copper (Cu), iron (Fe), manganese (Mn) and zinc (Zn), respectively while, the nutritional values in okra plants are presented accordingly in Table 2.

The elemental analysis for nutritional values of okra was carried out at the National Soil Plant, Fertilizer and water Laboratory, Umudike. Dry okra leaves were ground and milled to pass through 1 mm sieve. The ground samples were subjected to tissue analysis to determine content of Cu, Fe, Mn and Zn using the atomic absorption spectrometer (AAS).

2.6. Determination of growth components

Data on growth parameters were measured and recorded at 3, 5 and 7 weeks after planting (WAP), respectively. The okra plants flowered but, there were no fruits as most of the plant except for the control had almost shaded their leaves by 8WAP when the experiment was terminated. Several parameters were used in assessing the growth and productivity of the plant.

The plant height was determined from the soil level to the tip of the tallest leaf using a meter rule. The leaf number per plant was determined by counting, while the leaf area (LA) was determined by multiplying the length and width of the leaf. The measurements were taken fortnightly at 3WAP, 5WAP and 7WAP after the application of the SAR.

2.7. Statistical analysis

The data generated from the laboratory analysis and field experiment were subjected to statistical analysis of variance (ANOVA) and significant means were separated using Fishers' least significant different (FLSD) at 5% level of probability.

3. 0. Results and discussion

3.1. Effects of simulated acid rain and frequency of application on some phenological growth parameters of okra

Plant height: Figure 1 (A & B), shows the effects of SAR and frequency of SAR application on the plant height of *A. esculentus* @ 3, 5 and 7 WAS, respectively. Considering the effect of SAR (Fig. 1A), there was a significant decline in shoot heights from 3 to 7 WAS as concentrations decreased in pH value. The mean tallest plants were recorded at control (29.89 cm) followed by pH 4.5 with 26.70 cm and pH 2.7

produced the least plant height of 22.59 cm. Considering the effect of frequency of SAR application (Fig. 1B), there was equally significant reduction in shoot heights from 3 to 7 WAP as the frequency of SAR application increases from once to thrice a week. The mean tallest plant was recorded at an application of once a week (28.95 cm), while the mean shortest plant height of 17.47 cm was observed at an application of 3 times a week. With regards to the interaction effects (Table 1), higher plants were observed in the control pots with increase in growth from 3 to 7 WAP followed by a sequential reduction of the plants height as the concentrations decreases in pH value, as well as the increases in frequency of application from once weekly to thrice weekly, respectively.

From the results obtained, it was observed that the higher the acidity and frequency of application, the more the plant height of okra was inhibited. Reduction in plant height conforms to the observation of Tong and Liange, (2005) and is in agreement with the work of Dursun *et al.* (2002). The reduction in plant growth might have been caused by the injuries, reduced photosynthesis and other physiological disorders. In a similar study, Waldron (1978) stated that the pH level is responsible for reduction in plant growth. The observed stunted growth for plants treated with SAR pH 3.5 and 2.7 was similar to an earlier study by Singh and Agrawal (2001).

Leaf number: Figure 2 (A & B), shows the effect of SAR exposure at different pH levels and frequency of SAR application on leaf number of *A. esculentus*, respectively. The results indicated that under the stress of SAR and frequency of SAR application, number of leaves decreased with increased in acid rain, from 3 to 7 WAS. Considering the effect of SAR (Fig. 2A), the mean highest leaf number was determined at control (6.78) and while, the mean least leaf number of 3.63 was recorded in coke treatment. Considering the effect of frequency of SAR application (Fig. 2B), the mean highest leaf number (5.55) was determined at a weekly application of SAR while, the mean least number of leaf (2.75) was observed at an application of thrice a week. With regards to the interaction effects (Table 1), higher leaf numbers were observed in the control pots but, decrease in number with increase in plant growth from 3 to 7 WAS. This is in agreement with the work of Dursun *et al.*, (2002) and Tong and Liange, (2005) who confirmed that the higher the acidity, the more the number of leaves are inhibited.

Leaf area: Figure 3 (A & B), shows the effect of SAR exposure at different pH levels and frequency of SAR application on leaf area (LA) of *A. esculentus*, respectively. The results indicate that under the exposure of SAR and frequency of SAR application, the LA decreased with increased in acid rain, from 3 to 7 WAS. Considering the effect of SAR levels on the LA (Fig. 2A), the mean widest LA was determined at control (607.33 mm) and while, the mean least leaf number of 252.33 mm was recorded in coke treatment. Considering the effect of frequency of SAR application (Fig. 2B), the mean highest LA (436.67mm) was determined at a weekly application of SAR while, the mean least number of leaf (352.43 mm) was observed at an application of thrice a week. With regards to the interaction effects (Table 1), wider LA were observed in the control treatments but, decrease in

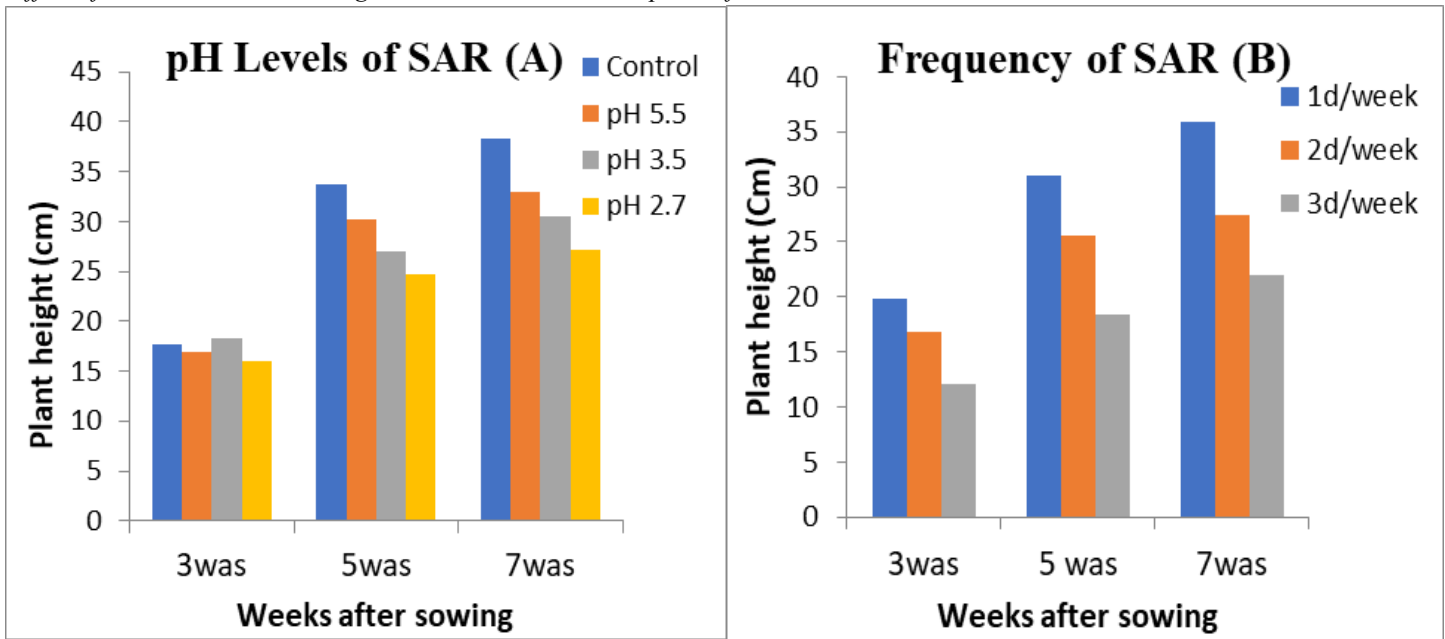


Figure 1: Effects of pH levels of SAR (A) and frequency of SAR (B) on plant height of okra @ 3, 5 and 7 weeks after sowing (WAS).

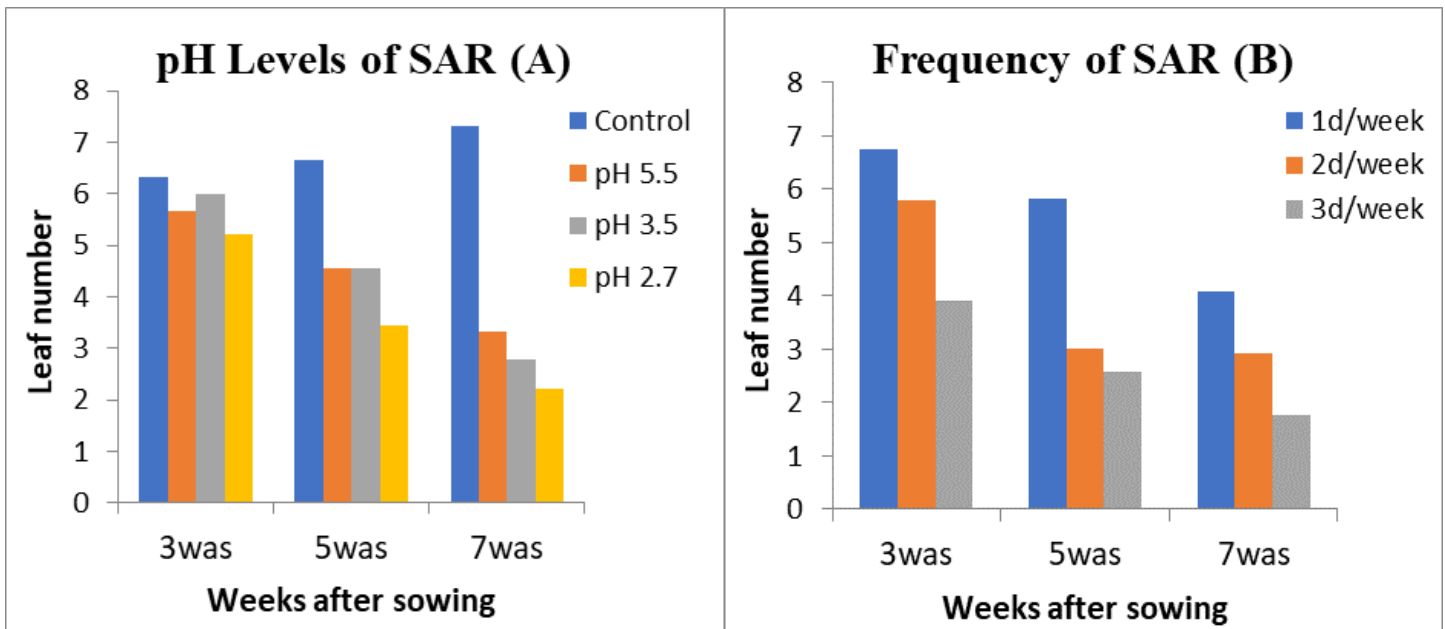


Figure 2: Effects of pH levels of SAR (A) and frequency of SAR (B) on plant leaf number of okra @ 3, 5 and 7 weeks after sowing (WAS).

leaf size with increase in plant growth from 3 to 7 WAS. The reduction in leaf size conforms to observation of (Tong and Liange, 2005).

The sequential reduction of leaf size of okra was proportional to the decrease in the concentration of pH value as well as, the increased in the frequency of SAR application to three times a week. This is in agreement with the work of Dursun *et al.*, (2002) and Tong and Liange, (2005), who confirmed that the higher the acidity, the more the LA was inhibition. The adverse effects of simulated acid rain on plant growth parameters on several crops were equally reported by Banwart *et al.* (1988), Evans *et al.* (1997) and Halman *et al.* (2008) respectively. A more recent report by Madiha *et al.* (2014) revealed that simulated AR also caused reduction in

plant growth and yield of field corn, green pepper and tomato.

The above results indicate that under the stress of simulated acid rain, the shoot height and number of leaves and size of leaf decreased with the declining pH value of acid rain, which affects the terminal buds of the plant. This is in agreement with the work of Dursun *et al.*, (2002). Size of the leaves also decrease due to thinner mesophyll cells. Reduction in leaf size conforms to observation of Liange (2005). Other morphological changes observed in this experiment (necrosis, chlorosis and deformation of leaf and terminal bud) is in accordance with the study of Dursun *et al.*, (2002) and Odiyi and Bamidele, (2013). It can be deduced from Table 1, that the control had the highest shoot height, number of leaves and size of leaf before and after exposure com-

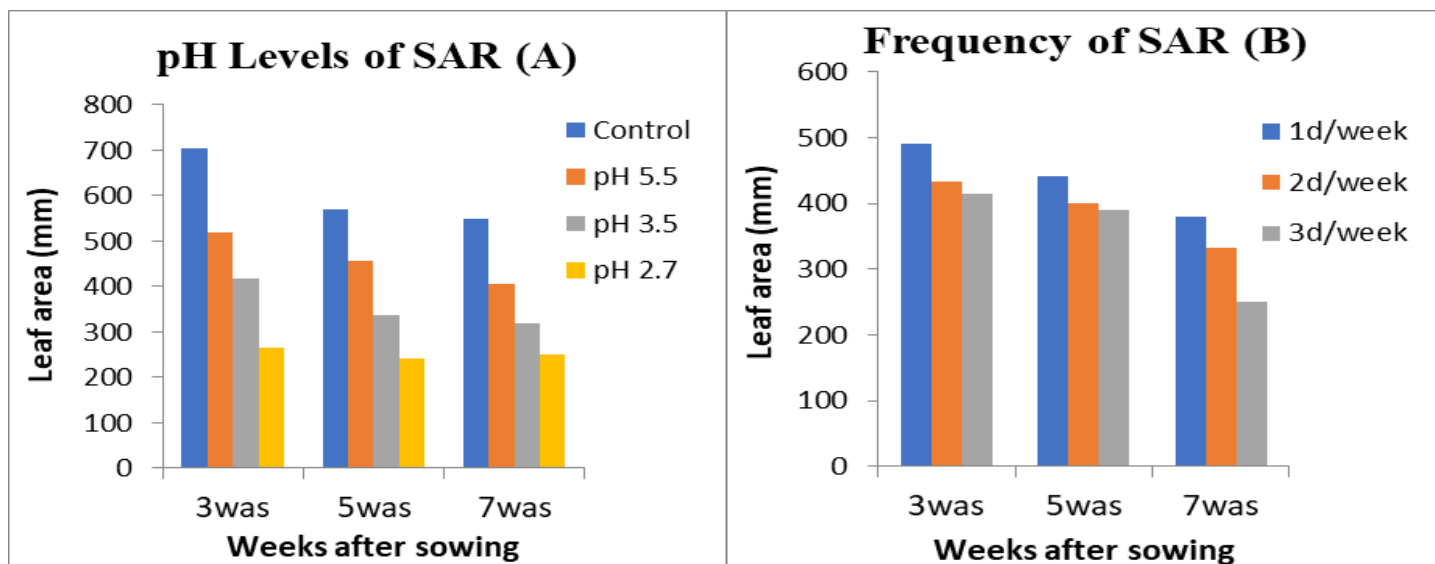


Figure 1: Effects of pH levels of SAR (A) and frequency of SAR (B) on plant height of okra @ 3, 5 and 7 weeks after sowing (WAS).

Table 1: Interaction effects of SAR and frequency of foliar spray on growth performance of Okra

Treatments	Plant height (cm)			Leaf number			Leaf area (cm ²)		
	3was	5was	7was	3was	5was	7was	3was	5was	7was
Control	21.33	34.33	38.33	7.34	6.17	5.53	703.0	571.00	548.0
Once weekly SAR application									
pH 5.5	18.67	33.67	37.67	6.33	5.40	4.03	626.0	549.0	503.0
pH 3.5	17.54	29.33	31.33	5.70	4.33	3.00	440.0	336.0	309.0
pH 2.7 (Coke)	16.00	24.33	27.33	4.70	3.53	2.37	328.0	282.0	277.0
Mean	17.43	29.11	32.11	5.58	4.42	3.13	465.0	389.0	363.0
Twice weekly SAR application									
pH 5.5	18.21	32.00	35.00	6.20	5.00	3.73	545.0	485.0	438.0
pH 3.5	17.00	27.33	31.00	5.40	4.30	2.87	389.0	334.0	289.0
pH 2.7 (Coke)	15.33	24.00	27.00	4.27	3.33	2.33	287.0	254.0	255.0
Mean	16.85	27.78	31.00	5.29	4.21	2.98	407.0	358.0	327.0
Thrice weekly SAR application									
pH 5.5	17.87	30.67	33.33	6.10	4.67	3.43	500.0	392.0	355.0
pH 3.5	16.67	25.33	27.93	5.30	4.13	2.67	310.0	306.0	273.0
pH 2.7 (Coke)	12.33	20.33	26.67	3.67	2.67	1.47	177.0	164.0	224.0
Mean	15.62	25.44	29.31	5.02	3.82	2.52	329.0	287.0	284.0
G. Mean	17.22	29.64	32.47	5.81	4.81	3.92	476.0	401.0	381.0
LSD (0.05)	5.11	6.04	7.76	1.90	1.57	1.15	293.1	227.6	188.4
CV (%)	17.50	12.00	14.10	19.30	19.30	17.40	36.40	33.50	29.20
F-test	N. S	N. S	N. S	N. S	N. S	N. S	N. S	N. S	N. S

Significance level of the F-test for interaction effects with NS denoting not significant, for two-sided t-test.

pared to pH 3.5 and 2.7 which had the least morphological parameters.

3.2 Effects of SAR and frequency of SAR on micronutrients (Cu, Fe, Mn and Zn) content in okra

The micronutrients content of *A. esculentus* were significantly ($P \leq 0.001$) affected by simulated acid rain (SAR) exposure at different pH levels (Table 2), frequency of SAR application (Table 2) and their interaction (Table 3), respectively. The results indicated that under the stress of simulated acid rain, micronutrients content in *A. esculentus* reduced with the declining pH value of acid rain and frequency of SAR application. Statistically, there was significant difference between SAR treatments and control plants.

Copper (Cu): There was a significant reduction in Cu content of the test plants as the concentrations of the SAR pH reduced in value. Considering the main effect of SAR (Table 2), there was significant increase in Cu content from pH 5.5 (4.11 mgkg⁻¹) to pH 2.7 (7.01 mgkg⁻¹) relative to control (3.27 mgkg⁻¹), with a mean Cu content of 4.98 mgkg⁻¹. Considering the effect of frequency of SAR application (Table 2), there was significant increase in Cu content from 1d/week (6.58 mgkg⁻¹) to 3d/week (11.63 mgkg⁻¹) with a mean Cu content of 9.08 mgkg⁻¹. With regards to the interaction effects (Table 3), higher Cu content (21.22 mgkg⁻¹) was determined in Coke (pH 2.7) treatment applied at 2d/week while, the lowest Cu content (14.42 mgkg⁻¹) was determined in pH 5.5 applied at 1d/weekly application of SAR relative

to control which were statistically not significant. The result is at par with previous studies reported by Arti *et al.*, 2010, Meng *et al.*, 2011, Eguagie, (2015).

Iron (Fe): Considering the main effect of SAR (Table 2) there was a significant increase in Fe content from 51.77 mgkg⁻¹ (pH 5.5) to 93.42 mgkg⁻¹ (pH 2.7), relative to control (34.31 mgkg⁻¹) with a mean Fe content of 64.86 mgkg⁻¹. Considering the effect of frequency of SAR application (Table 2), there was significant increase in Fe content from 49.94 mgkg⁻¹ (1d/week) to 88.99 mgkg⁻¹ (3d/week) with a mean Fe content of 69.85 mgkg⁻¹. With regards to the interaction effects (Table 3), higher Fe content (206.03 mgkg⁻¹) was determined in Coke (pH 2.7) solution applied at 3d/week while, the lowest Fe content of 63.34 mgkg⁻¹ was determined in SAR (pH 5.5) applied at 1d/week relative to the control. The result is at par with previous studies reported by Arti *et al.*, 2010, Meng *et al.*, 2011, Eguagie, (2015).

Manganese (Mn): Considering the main effect of SAR (Table 2), the mean highest Mn content (85.63 mg kg⁻¹) was determined under Coke (pH 2.7) while, mean lowest Mn value (49.43 mgkg⁻¹) in okra was obtained under pH 5.5 treatment relative to control (22.91 mgkg⁻¹) with an average Mn content of 55.11 mgkg⁻¹. Considering the effect of frequency of SAR application (Table 2), there was significant increase in Mn content from 1d/week (37.91 mgkg⁻¹) to 3d/week (91.27 mgkg⁻¹) with a mean Mn content of 55.40 mgkg⁻¹. With regards to the interaction effects (Table 4.5), higher Mn content of 395.51 mgkg⁻¹ was determined in Coke (pH 2.7) treatment applied at 3d/week while, the lowest Mn content of 123.66 mgkg⁻¹ was determined in pH 5.5 applied at 1d/week relative to the control. The result is at par with previous stud-

ies reported by Wang *et al.*, (2000), Khan and Shikha (2004), Khan and Deopura (2005), Meng *et al.*, 2011, Imran *et al.*, (2014).

Zinc (Zn): Considering the main effect of SAR (Table 2), the mean highest Zn content (68.84 mg kg⁻¹) was determined under Coke (pH 2.7) solution while, mean least Zn value (32.16 mgkg⁻¹) in okra was obtained under pH 5.5 treatment relative to the control (15.73 mgkg⁻¹) with a mean value of 41.98 mgkg⁻¹. Considering the effect of frequency of SAR application (Table 2), there was significant increase in Zn content from 1d/week (27.02 mg kg⁻¹) to 3d/week (63.77 mgkg⁻¹) with a mean Mn content of 45.08 mg kg⁻¹. With regards to the interaction effects (Table 4.5), higher Zn content (142.02 mgkg⁻¹) was determined in Coke (pH 2.7) treatment applied at 2d/week while, the lowest Zn content (34.84 mg kg⁻¹) was determined in pH 5.5 applied at 1d/weekly application of SAR. The result is at agreement with previous studies reported by Meng *et al.*, (2011) and Eguagie *et al.*, (2016).

4.0. Conclusion

This study examined the impact of simulated acid rain and frequency of SAR on growth and micronutrients content of okra (*Abelmoschus esculentus*) in an ultisol at Umudike.

It is evident from this study that under the stress of simulated acid rain (SAR) exposure at pH value below 5.5, had negative effect on the growth component (shoot height, number of leaves and LA) which decreased with the declining pH value of acid rain and this markedly suppressed fruit formation of *A. esculentus* due to the reduction of photosynthesis as a result of chlorosis, necrosis and leaf abscission coupled with dehydration of the plants.

Table 2: Effects of SAR and frequency of SAR application Cu, Fe, Mn and Zn content in okra

Treatments	Micronutrients (mgkg ⁻¹) in okra leaf			
	Cu	Fe	Mn	Zn
pH levels of SAR				
Control	3.27	34.31	22.91	15.73
pH 5.5	4.11	51.77	49.43	32.16
pH 3.5	5.54	79.95	62.45	51.17
pH 2.7 (coke)	7.11	93.42	85.63	68.84
Mean	5.01	64.86	55.11	41.98
LSD (0.05)	1.97	42.44	55.99	6.44
CV (%)	22.21	25.60	18.42	15.81
F-test	***	***	***	***
Frequency SAR application				
1d/weekly	6.58	49.94	37.91	27.02
2d/weekly	9.04	70.63	57.03	44.45
3d/weekly	11.63	88.99	91.27	63.77
Mean	9.08	69.85	55.40	45.08
LSD (0.05)	1.71	36.76	48.49	5.58
CV (%)	22.2	25.60	18.44	15.82
F-test	***	***	***	***

Significance level of the F-test for treatment effects with *** denoting P < 0.001, for two-sided t-test.

Table 3: Interaction effects of SAR and frequency of SAR on Micronutrients content of Okra

Treatments	Micronutrients (mgkg ⁻¹)			
	Cu	Fe	Mn	Zn
Control	12.50	43.43	106.57	29.37
1d/weekly				
pH 5.5	14.42	63.43	124.66	34.84
pH 3.5	15.50	91.35	221.34	44.03
pH 2.7 (coke)	18.63	115.01	312.07	67.02
Mean	16.18	89.93	219.36	48.63
2d/weekly				
pH 5.5	16.30	97.35	149.72	42.61
pH 3.5	19.50	108.06	267.32	57.22
pH 2.7 (coke)	21.22	125.47	345.39	89.55
Mean	19.01	110.29	254.14	63.13
3d/weekly				
pH 5.5	19.83	114.76	171.16	69.27
pH 3.5	15.17	172.22	288.56	66.88
pH 2.7 (coke)	10.33	206.03	395.51	142.02
Mean	15.11	164.33	285.08	92.72
G. Mean	13.11	121.52	271.51	90.75
LSD (0.05)	3.42	73.52	96.97	11.16
CV (%)	22.20	25.60	18.43	15.82
F-test	N. S	***	**	**

Significance level of the F-test for interaction effects with NS, *, ** and *** denoting not significant, $P < 0.05$, 0.01 and 0.001, respectively, for two-sided t-test.

Except for the accumulation of micronutrients, which increased with declining pH value of acid rain and frequency of application, all the plant growth parameters studied such as plant height, number of leaves and leaf area of *A. esculentus* decreased significantly with the declining pH value of acid rain and frequency of application with respect to the control treatment.

Coke solution (pH 2.7 levels) and three (3) days per weekly application of SAR gave the highest reduction in plant growth parameter in Okra but, had higher increased in micronutrients, respectively.

References

- Arti, V., Ashish, T. and Abdullah, A. (2010). An impact of simulated acid rain of different pH levels on some major vegetable plants in India. Report and Opinion. 2(4):38-40.
- Banwart, W. L., Porter, P. M., Ziegler, E. L. and Hassett, J. J. (1988). Growth parameters and yield component response of field corn to simulated acid rain. Environ. Expt. Bot. 28:43-51. Available: [http://dx.doi.org/10.1016/0098-8472\(88\)90045-7](http://dx.doi.org/10.1016/0098-8472(88)90045-7).
- Butnariu, M. and Samfira, I. (2013). Consequences of Acid Rain. *Journal of Ecosystem and Ecography*, Vol. 4 (2): 1-3
- Chesapeake B. (2013). Ecological Foundation, Inc. Acid Rain Overview. Available: <http://www.chesbay.org/acidRain/>
- Dubay, D.T. and Allen, S.H. (1987). The effects of simulated acid rain with and without ambient rain on the growth and yield of field-grown soybeans. Environ. Exp. Bot; 27(4): 395.
- Dursun, A., Kumlay, A.M., Yilderin. E. and Guvenç, I. (2002). Effects of simulated acid rain on plant growth and yield of tomato. Acta Hort. 579:245-248.
- Eguagie, M. O. (2015). Effects of simulated acid rain on the growth, yield and mineral nutrient relations of *Solanum lycopersicum* L. European Journal of Biotechnology and Bioscience. 3(11):15-18.
- Eguagie, M. O., Aiwansoba, R. P., Omofomwan, K. O. and Oyanoghafo, O. O. (2016). Impact of Simulated Acid Rain on the Growth, Yield and Plant Component of *Abelmoschus caillei*. *Journal of Advances in Biology & Biotechnology*, 6(1): 1-6.
- Eteng, E. U., Asawalam, D. O. and Ano, A. O. (2014). Effect of Cu and Zn on maize (*Zea mays* L.) yield and nutrient uptake in coastal plain sand derived soils of southeastern Nigeria. *Open Journal of Soil Science (OJSS); Journal Of Scientific Research Publishing*. Vol. 4 (7): 235-245).
- Evans, L.S., Gmor, N. F. and Dacosta, F. (1997). Leaf surface and histological perturbations of leaves of *Phaseolus vulgaris* and *Helianthus annuus* after exposure to simulated acid rain. American Journal of Bot. 4:304-313.
- Huang, X. H., Zeng, O.L. and Zhou, Q. (2005). Effect of acid rain on seed germination of rice, wheat and grape. Huan Jing Kexue. 26:181-184.
- IITA, 1979. Selected methods for soil and plant analysis. International Institute of Tropical Agriculture Manual Series No. 1, Ibadan, p. 62.
- Imran, M. A. Hussain, S., Hussain, M., Nawaz Ch, M. and Meo, A. A. (2014). Effect of simulated acid rain (sar) on some morphochemical aspects of mash (*Vigna mungo* L.) *Pak. J. Bot.*, 46(1): 245-250.

- Johnston, J.W., Shriner, D.S. and Kinerley, C. K. (1986). The combined effects of simulated acid rain and ozone on injury, chlorophyll and growth of radish. *Environmental and Experimental Botany*; 26(2): 107.
- Kausar, S., Khan, A.A. and Rahman, F. (2005). Impact of simulated acid rain on potato. In herbal drug and environmental pollution. A Satellite Session of Third International Conference on Plants and Environmental Pollution, Jamia Hamdard, New Delhi.
- Khan, T. I., and Deopura, S. (2005). Physiological and biochemical effects of simulated acid rain on *Phaseolus vulgaris* Var HUR 15. *The Environmentalist*, 24, 224-226.
- Khan, T.I. and Shikha, D. (2004). Physiological and biochemical effects of simulated acid rain on *Phaseolus vulgaris* var. HUR-15. *Environmentalist*; 24(4): 223.
- Kita, I., Sato, T., Kase, Y. and Mitropoulos, P. (2004). Neutral rains at Athens, Greece: a natural safeguard against acidification of rains. *Sci Total Environ.*, 327:285-294.
- Lal, N. and Singh, H. (2012). The effects of simulated acid rain of different pH-levels on biomass and leaf area in Sunflower (*Helianthus annuus*). *Current Botany*; 2012, 3(5): 2220-4822.
- Li, W. and Gao. J. (2002). Acid deposition and integrated zoning control in China. *Environ. Manag.*, 30(2): 169-182.
- Madiha, Y. H., Abrar, A.K. and Zubairu, U.D. (2014). Effect of acid rain on growth of Pawpaw (*Carica papaya*) and Castor (*Ricinus communis*) plants. *Journal of Science and Technology*; 1(9): 43-48.
- Meng, H. Dong, D., Wang, J., Yang, K., Tian, L., Sun, W. and Fang, C. (2011). Effects of simulated acid rain on main nutritional indicators of three leafy vegetables. *Chemical Research Chinese University*; 27(3): 397-401.
- Milton, M. B. and Abigael, T. O. (2015). Effects of simulated acid rain on the morphology, phenology and yield of Okra (*Abelmoschus esculentus* (L). Moench) *Journal of Environmental Science, Computer Science and Engineering & Technology Sec. A*; Vol.4. No. 2, 501-518.
- Nduka, J.K.C., Oriskwe, O.E., Ezenweke, L.O., Ezenwa, T.E., Chendo, M.N. and Ezeabasili, N.G. (2008). Acid rain phenomenon in Niger Delta region of Nigeria: economic, biodiversity and public health concern. *The SciWorld J* 8: 811-818
- NRCRI (2018). Federal Ministry of Agriculture and Rural Development. National Root Crops Research Institute, Umudike. Annual Report, 2018.
- Odiyi, B. O. and Bamidele, J. F. (2014). Effects of simulated acid rain on growth and yield of Cassava *Manihot esculenta* (Crantz). *Journal of Agricultural Science*. 6(1): 96-101.
- Odiyi, B. O. and Eniola, A. O. (2014). The Effect of simulated acid rain on plant growth component of Cowpea (*Vigna unguiculata*) L. Walps. *Jordan Journal of Biological Sciences*. 2014;8(1):51-54.
- Prashant, M. P. (2010). Science behind Acid Rain: Analysis of Its Impacts and Advantages on Life and Heritage Structures. *South Asian Journal of Tourism and Heritage*. Vol. 3, No. 2 [accessed May 12 2019]
- Sant'Anna-Santos, B. F., Silva, L. C., Azevedo, A. A. and Deque-Brasil, R. (2006). Effects of simulated acid rain on photosynthesis, Chlorophyll fluorescence and antioxidative enzymes in *Cucumis sativus* L. *Photosynthetica*; 40: 331-335.
- Singh, A. and Agrawal, M. (2008). Acid rain and its ecological consequences. *Journal of Environmental Biology*, 29 (1), 15-24.
- Singh, B. and M. Agrawal (2004). Impact of simulated acid rain on growth and yield of two cultivars of wheat, *Water Air Soil Pollut.*, vol. 152, pp. 1-4.
- Singh, R.K. and Agrawal, M. (2001). Effect of industrial emission on atmospheric wet deposition. *Water Air Soil Pollution*; 130: 481-486.
- Stoynora, D., and Velilova, V. (1998). Effects of simulated acid rain on chloroplast ultrastructure of primary levels of *Phaseolus vulgaris*. *Biological plant*, 40, 589-598.
- Sunita, B. S. and Sharad, B. S. (2013). Ecological consequences of the Acid rain. *IOSR Journal of Applied Chemistry (IOSR-JAC) Volume 5, Issue 4:19-24*.
- Udo, E. J. Ibia, T. O. Ogunwale, J. A., Ano, A. O. and Esu, I. E. 2009. Manual of Soil, Plant and Water Analysis. Sibon Books Limited, Lagos.
- Tong, G. and Liange, H. (2005). Effect of simulated acid rain and its acidified soil on soluble sugar and nitrogen contents of wheat seedlings. *Xin Young Sheng Tai Xue Bao*. 16: 1487-1492.
- Velikova, V. K., Yordanov, K.M. and Tsonev, T. (1997). Effects of simulated acid rain on the photosynthetic characteristics of *Phaseolus vulgaris* L. *Photosynthetica*; 34: 523-535.
- Wang, H. F., Takematsu, N. and Ambe, S. (2000). Effects of soil acidity on the uptake of trace elements in soybean and tomato plants, *Appl. Rad. Isot.*, 52, 803-811.